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High Prevalence Fluke Infection at Four Cattle Farms Located in Kuala Terengganu, Malaysia

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Abstrak: Satu kajian telah dijalankan untuk mengenal pasti kehadiran cacing parasit pada lembu di empat ladang di Kampung Surau Haji Daud, Kampung Kubang Jela, Kampung Gemuruh dan Kampung Peradong, Kuala Terengganu. Sebanyak 33 sampel tinja dan darah telah diambil daripada lembu yang terlibat dalam kajian ini. Sampel tinja diproses dengan kaedah pemendapan, kaedah McMaster yang telah diubah suai, dan kultur tinja, untuk mengesan telur trematod, nematod dan juga untuk mengenal pasti larva peringkat ketiga. Sampel darah diproses dengan kaedah *Packed Cell Volume* (PCV) untuk mengetahui sama ada lembu adalah anemik, normal atau mengalami dehidrasi. Keputusan menunjukkan bahawa daripada 33 ekor lembu yang diperiksa, 17% daripadanya adalah positif untuk cacing fluk hati, 67% adalah positif untuk cacing fluk perut, dan 42% adalah positif untuk jangkitan nematod. Nilai PCV menunjukkan bahawa tiada lembu yang anemik atau mengalami dehidrasi. Keputusan telah dihantar ke Jabatan Perkhidmatan Veterinar untuk rawatan dan tindakan seterusnya.

Kata kunci: Paramphistomum, Fasciola, Lembu, Malaysia

Abstract: A survey was conducted to determine the presence of parasitic worm infection in cattle at four farms located in Kampung Surau Haji Daud, Kampung Kubang Jela, Kampung Gemuruh and Kampung Peradong, Kuala Terengganu. Thirty-three faecal samples and blood samples were obtained from each cattle involved in this survey. Faecal samples were subjected to sedimentation method, modified McMaster method and faecal culture to detect trematode eggs, nematode eggs, and to identify the third stage of larvae. Blood samples were subjected to Packed Cell Volume (PCV) method to determine if the cattle are anaemic,

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normal or dehydrated. Result indicated that out of 33 cattle examined 17% of the cattle were positive for liver fluke, 67% were positive for stomach fluke and 42% were positive for nematode infection. PCV value indicated that all the cattle examined are not anaemic or dehydrated. The results obtained showed that trematode and nematode infections are common problem in all the four farms. The results were submitted to Department of Veterinary Services to plan further action and treatment.

Keywords: Paramphistomum, Fasciola, Cattle, Malaysia

INTRODUCTION

Parasites infection is one of the most important causes of the production losses of meat and milk in most cattle-producing countries of the world (Chiu *et al.* 2014). The economic impact of gastrointestinal parasite infection results in mortality losses and retarded in growth and production of food (Rajakaruna & Warnakulasooriya 2011). Effects on animal productivity are common following parasite infection, affecting weight gain and milk production. Clinical effects following parasite infections are anaemia, oedema and diarrhoea (Bowman 2014).

There were 90,480 cattle recorded in Terengganu in 2014 (Department of Veterinary Services 2017). Parasitic infection may affect the health and productivity of cattle which eventually leads to considerable economic losses. Several studies (Khadijah *et al.* 2015; 2017; Rita *et al.* 2017) on parasitic infection in cattle have been carried out in Terengganu, Malaysia. This survey was conducted with the aim to determine the current prevalence of parasitic worm infection in Kuala Terengganu, specifically stomach fluke and liver fluke infection, in order to add more information on the current status of parasitic infection of cattle in Kuala Terengganu.

MATERIALS AND METHODS

Farms Location and Information

The survey was performed at four cattle farms in Kuala Terengganu, namely Kampung Surau Hj Daud (Farm A), Kampung Kubang Jela (Farm B), Kampung Gemuruh (Farm C) and Kampung Peradong (Farm D), Kuala Terengganu (Fig. 1). Animals at all the farms were allowed to graze in the morning until late afternoon in open pastures. For Farm D, the animals were also allowed to graze under rubber trees.

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Figure 1: Location of sampling sites. Source: Google Earth and GIS version 2.14.1

Animals

Thirty-three cattle were selected in this survey and the breed involved was Kedah-Kelantan. The cattle were aged between 1.5 to 5 years old and at each farm, at least 30% of the cattle were randomly sampled.

Sample Collections

Faeces

Faeces were collected individually from the rectum of the cattle following the guidelines by Ministry of Agriculture, Fisheries and Food (1986). These faecal samples were brought back to the laboratory and were kept at 4°C until processing for different tests. The samples were subjected to tests for trematode egg count using sedimentation method (Ministry of Agriculture, Fisheries and Food 1986) nematode worm egg count using McMaster method (Ministry of Agriculture, Fisheries and Food 1986) and faecal culture (Ministry of Agriculture, Fisheries and Food 1986) for third stage larvae identification of nematodes (Ministry of Agriculture, Fisheries and Food 1986).

Blood

Blood samples were withdrawn from the cattle (Kerr 2002) either from the tail vein or the jugular vein using needles sized 18 G into 3 ml blood tubes with Ethylenediamine-tetraacetic acid (EDTA) anticoagulant (Vacutest Kima brand, Italy). The

whole blood was subjected to the Packed Cell Volume method (Rosenberger *et al.* 1979) to determine the proportion of erythrocytes in blood.

Examination of Parasites and Packed Cell Volume (PCV)

Trematodes

Sedimentation method (Ministry of Agriculture, Fisheries and Food 1986) was conducted to determine the trematode egg count in faecal samples. The number of eggs observed in the sediment was divided by 5 to get the value of eggs per gram faeces.

Nematodes

Modified McMaster Method was performed for the estimation of nematode eggs in 1 g of faeces according to Ministry of Agriculture, Fisheries and Food (1986). The number of eggs observed was multiplied by 100 to get the value of eggs per gram faeces.

Faecal culture method was conducted to obtain the third stage larvae (L_3) from faecal samples. The L_3 was identified based on the identification keys provided by Ministry of Agriculture, Fisheries and Food (1986).

Packed Cell Volume (PCV)

PCV was conducted according to Rosenberger *et al.* (1979). The PCV values obtained were recorded and then differentiated with the normal blood values of cattle in order to determine if they were either anaemic, normal or dehydrated.

Statistical Analysis

Normality test was conducted in order to know the distribution of data (number of worm egg). The data was not normally distributed. Thus, Kruskal-Wallis test was conducted to compare the WEC (for nematodes and trematodes) between four different farms. Chi-square test was performed to differentiate the frequency of third stage larvae present at the farms. Statistical analyses were performed using SPSS version 22 (IBM Corporation, USA).

RESULTS

Prevalence of Trematode Infection

From 33 samples, 6 (18%) were found positive for liver flukes and 27 samples (81%) were found positive for stomach fluke.

Fasciola and Paramphistomum worm egg counts do not differ across the four farms, $\chi^2(3, N = 33) = 1.269$, p > 0.05 and $\chi^2(3, N = 33) = 4.950$, p > 0.05 respectively (Table 1).

Egg count (e.p.g) Number of Farm location Fasciola Paramphistomum sp. animal sp. Kampung Surau Hj Daud 9 22.2 ± 44.1 1440 ± 1810.6 Kampung Kubang Jela 9 22.2 ± 44.1 244.4 ± 212.8 Kampung Gemuruh 10 80 ± 168.7 2110 ± 2373.2 0 Kampung Peradong 5 1560 ± 1128.3 Kruskal-Wallis 1.269 4.950 0.737 0.176 p-value

Table 1: Number of egg count (mean ± standard deviation) for liver fluke (*Fasciola* sp.) eggs and stomach fluke eggs (*Paramphistomum* sp.) in cattle at four farms in Kuala Terengganu.

Prevalence of Nematode Infection

The nematode egg count for all four farms ranged between 0–1000 e.p.g. The highest nematode egg count was recorded in Kampung Kubang Jela with 1000 e.p.g while the rest of the farms were recorded with lowest nematode egg count of 0 e.p.g. There was no significant different of nematode egg count distribution between the four farms, $\chi^2(3, N = 33) = 6.782$, p > 0.05 (Table 2).

 Table 2: Mean worm egg count (mean ± standard deviation) of nematode eggs in cattle at four different farms in Kuala Terengganu.

Farm location	Number of animals	Mean worm egg count (e.p.g)
Kampung Surau Hj Daud	9	100 ± 100.0
Kampung Kubang Jela	9	275 ± 369.9
Kampung Gemuruh	10	20 ± 42.0
Kampung Peradong	5	100 ± 141.4
Kruskal-Wallis		6.782
<i>p</i> -value		0.079

Third Stage Larvae (L₃) Identification

The most dominant nematode recorded was *Haemonchus* spp., followed by *Oesophagostomum* sp. There was no significant difference between the numbers of *Haemonchus* spp. on the four farms $\chi^2(3, N = 393) = 0.374$, *p* > 0.05 (Table 3).

Table 3: The percentage of *Haemonchus* and *Oesophagostomum* L_3 in cattle at four different farms in Kuala Terengganu.

Farm location	Percentage (%)	
	Haemonchus spp.	Oesophagostomum sp.
Kampung Surau Hj Daud	100	0
Kampung Kubang Jela	100	0
Kampung Gemuruh	93	7
Kampung Peradong	100	0
Chi-square	0.374	
<i>p</i> -value	0.946	

Packed Cell Volume

The PCV recorded ranged between 24%–48% for all the cattle sampled.

DISCUSSION

This present survey recorded high prevalence of trematode and low prevalence of nematode infection in cattle at four farms in Kuala Terengganu, Malaysia. All four farms reported the presence of stomach fluke while three farms reported the presence of liver fluke.

The prevalence of stomach fluke infection in this current survey was high (81%) compared to liver fluke infection. It is also reported to be higher than the findings of Khadijah *et al.* (2017) and Rita *et al.* (2017) which recorded the percentage of stomach fluke infection in cattle Terengganu, Malaysia as 15% and 59% respectively. The report of stomach fluke incidences coincides with other reports which suggested that stomach fluke is highly reported in areas with sub-tropical and tropical climate, such as in Asia, Africa and Russia (Horak 1971; Gupta *et al.* 1978). However, studies on stomach fluke infection in Terengganu and Malaysia are very limited. Therefore, more studies on prevalence of stomach fluke infection in Malaysia and the epidemiology are suggested, due to the fact that this parasite was reported to cause production losses in cattle and buffalo (Saleha 1991).

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The prevalence of liver fluke infection in this present survey was lower (18%) than the those reported by Khadijah *et al.* (2015; 2017) and Rita *et al.* (2017) where the prevalence were recorded as 94.6% and 67.8%, respectively. However, the infection of liver flukes in this present survey was supported by previous researchers who reported that liver fluke infection affected cattle (Saleha 1991; Rajamanickam *et al.* 1996) and buffaloes in Malaysia (Khadijah *et al.* 2017; Rosilawati & Saipul Bahari 2017). Similar to stomach fluke studies, studies on liver fluke infection in Terengganu and Malaysia are very limited, with very few literatures published after the findings of Rajamanickam *et al.* (1996). Therefore, more studies on liver fluke infection in Malaysia and the epidemiology are suggested, due to the fact that this parasite was reported to cause production losses in cattle, sheep and goats (Saleha 1991).

Haemonchus spp. was found to be highly prevalent followed by *Oesophagostomum* sp. This is supported by Waruiru *et al.* (2001) who reported that *Haemonchus* spp. were recorded as the predominant species among the trichostrongylids in cattle.

Despite having *Haemonchus* spp. in abomasum and infected with flukes, percentage of PCV in all the animals were ranged between 24%–48%, indicating normal range as reported by Rajamanickam *et al.* (1987). This finding was not expected as the animals were infected by blood-sucking parasites, and PCV values should be significantly lower as reported by Rajamanickam *et al.* (1987). PCV might not be affected by *Haemonchus* spp. infection as the infection was low, with mean worm egg counts were 100 and 275 e.p.g. Similar to this, mean Fasciola egg counts were 22 and 80 per gram, indicating low infection that might not be affecting PCV values. On the other hand, it was expected that infection by Paramphistomum with mean worm egg count ranged between 244–2110 e.p.g. will affect the PCV value. However, this is not the case in this current survey. It was reported that the immature stages suck blood and are responsible for anaemia in cattle (Diaz *et al.* 2006) and it is probable that in this current survey the immature stages are low when compared to adults which produce eggs.

It is suggested that in future studies, observation and sampling of freshwater snails could be conducted to determine the source of fluke infections as in this current study the snails were not observed. Information on the availability of the snails as source of infection will help farmers to manage and control fluke infection.

CONCLUSION

Present survey recorded high prevalence of trematode infection and low prevalence of nematode infection. This survey revealed that the animals were infected with liver fluke, stomach fluke, *Haemonchus* spp. and *Oesophagostomum* sp. Although, the PCV value showed that all of the cattle sampled were in healthy condition, regular monitoring by Department of Veterinary Services is required. Similar survey with wider coverage of the farms in Terengganu and other states should be conducted

to provide more information on trematode and nematode infection that will assist farmers and Department of Veterinary Services to improve animal's health.

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REFERENCES

- Bowman D D. (2014). *Georgis' parasitology for veterinarians* (10th Ed.). USA: Elsevier Inc, 496.
- Chiu C H, Lian C W, Chien H P, Cheng H Y and Cheng H L. (2014). Investigation of gastrointestinal parasites of dairy cattle around Taiwan. *Journal of Microbiology, Immunology and Infection* 47(1): 70–74. https://doi.org/10.1016/j.jmii.2012.10.004
- Department of Veterinary Services Malaysia. (2017). Number of livestock 2015. http://www. dvs.gov.my/en/statistic [Accessed on 12 September 2017].
- Díaz P, Lomba C, Pedreira J, Arias M, Sánchez.Andrade R, Suárez J L, Díez-Baños P, Morrondo P and Paz-Silva A. (2006). Analysis of the IgG antibody response against Paramphistomidae trematoda in naturally infected cattle. Application to serological surveys. *Veterinary Parasitology* 140: 281–288. https://doi.org/10.1016/j. vetpar.2006.04.007
- Gupta P P, Singh B and Dutt S C. (1978). A note on amphistomiasis in an adult buffalo. *Indian Veterinary Journal* 55: 491–492.
- Horak I G. (1971). Paramphistomiasis of domestic ruminants. *Advance in Parasitology* 9: 33–71. https://doi.org/10.1016/S0065-308X(08)60159-1
- Kerr M G. (2002). Veterinary laboratory medicine: Clinical biochemistry and haematology (2nd Ed.). Oxford: Blackwell Science Ltd., 368.
- Khadijah S, Izzudin A H, Rita N, Veronica S, Aida H and Wahab A R. (2015). Endo-and ectoparasite infections in two cattle farms located in Kuala Terengganu, Peninsular Malaysia. *Asian Journal of Agricultural and Food Sciences* 3: 667–674.
- Khadijah S, Ariff Z, Nurlaili A, Sakiinah A, Izzudin A H, Mursyidah A K, Rita N and Nur Aida H. (2017). Fasciola and paramphistomum infection in large ruminants. *International Journal of Agronomy and Agricultural Research* 10(6): 19–26.
- Ministry of Agriculture, Fisheries and Food. (1986). *Manual of veterinary parasitological laboratory techniques*. London: Her Majesty's Stationary Office, 160.
- Rajakaruna R S and Warnakulasooriya K N. (2011). Gastrointestinal parasites in dairy cattle in kandy district in Sri Lanka study area collection of data and samples. *Annual Research Journal of Sri Lanka Students' Association in Japan* 11: 92–99.
- Rajamanickam C, Cheah T S, Chandrawathani P, Hasnah Y and Adnan M. (1996). *Fascioliasis in Peninsular Malaysia*. Putrajaya: Department of Veterinary Services, Ministry of Agriculture Malaysia.

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- Rajamanickam C, Muniandy M, Yahya M, Cheah T S and Paramasvaran S. (1987) Haematological and biochemical parameters of cattle naturally infested with *Fasciola gigantica* in Perak, Malaysia. *Kajian Veterinar* 19(1): 143–151.
- Rita N, Mursyidah A K and Khadijah S. (2017). The prevalence of helminthiasis in cattle, Terengganu, Peninsular Malaysia. *Tropical Biomedicine* 34(2): 324–331.

Rosenberger G, Dirksen G, Gründer H D, Grunert E, Krause D and Stöber M. (1979). *Clinical examination of cattle*. Berlin: Verlag Paul Parey, Berlin and Hamburg.

- Rosilawati K and Saipul Bahari A R. (2017). Fascioliasis in and adult draught buffalo in Malaysia: A case report. *Malaysian Journal of Veterinary Research* 8(1):1 69–172.
- Saleha A A. (1991). Liver fluke disease (fascioliasis): Epidemiology, economic impact and public health significance. *The Southeast Asian Journal of Tropical Medicine and Public Health* 22(Supplement): 361–364.
- Waruiru R M, Thamsborg S M, Nansen P, Kyvsgaard C N, Bogh H O, Munyua W K and Gathuma J M. (2001). The epidemiology of gastrointestinal nematodes of dairy cattle in central Kenya. *Tropical Animal Health and Production* 33: 173–187.